

RESEARCH ARTICLE SUMMARY

STRUCTURAL BIOLOGY

Substrate-engaged 26S proteasome structures reveal mechanisms for ATP-hydrolysis-driven translocation

Andres H. de la Peña*, Ellen A. Goodall*, Stephanie N. Gates*, Gabriel C. Lander†, Andreas Martin†

INTRODUCTION: As the major protease in eukaryotic cells and the final component of the ubiquitin-proteasome system, the 26S proteasome is responsible for protein homeostasis and the regulation of numerous vital processes. Misfolded, damaged, or obsolete regulatory proteins are marked for degradation by the attachment of polyubiquitin chains, which bind to ubiquitin receptors of the proteasome. A heterohexameric ring of AAA+ (ATPases associated with diverse cellular activities) subunits then uses conserved pore loops to engage, mechanically unfold, and translocate protein substrates into a proteolytic core for cleavage while the deubiquitinase Rpn11 removes substrate-attached ubiquitin chains.

RATIONALE: Despite numerous structural and functional studies, the mechanisms by

which adenosine triphosphate (ATP) hydrolysis drives the conformational changes responsible for protein degradation remained elusive. Structures of related homohexameric AAA+ motors, in which bound substrates were stabilized with ATP analogs or hydrolysis-eliminating mutations, revealed snapshots of ATPase subunits in different nucleotide states and spiral-staircase arrangements of pore loops around the substrate. These structures gave rise to “hand-over-hand” translocation models by inferring how individual subunits may progress through various substrate-binding conformations. However, the coordination of ATP-hydrolysis steps and their mechanochemical coupling to propelling substrate were unknown.

RESULTS: We present the cryo-electron microscopy (cryo-EM) structures of the actively

ATP-hydrolyzing, substrate-engaged 26S proteasome with four distinct motor conformations. Stalling substrate translocation at a defined position by inhibiting deubiquitination led to trapped states in which the substrate-attached ubiquitin remains functionally bound to the Rpn11 deubiquitinase, and the scissile isopeptide bond of ubiquitin is aligned with the substrate-translocation trajectory through the AAA+ motor. Our structures suggest a ubiquitin capture mechanism, in which mechanical pulling on the substrate by the AAA+ motor delivers ubiquitin modifications directly into the Rpn11 catalytic groove and accelerates isopeptide cleavage for efficient, cotranslocational deubiquitination.

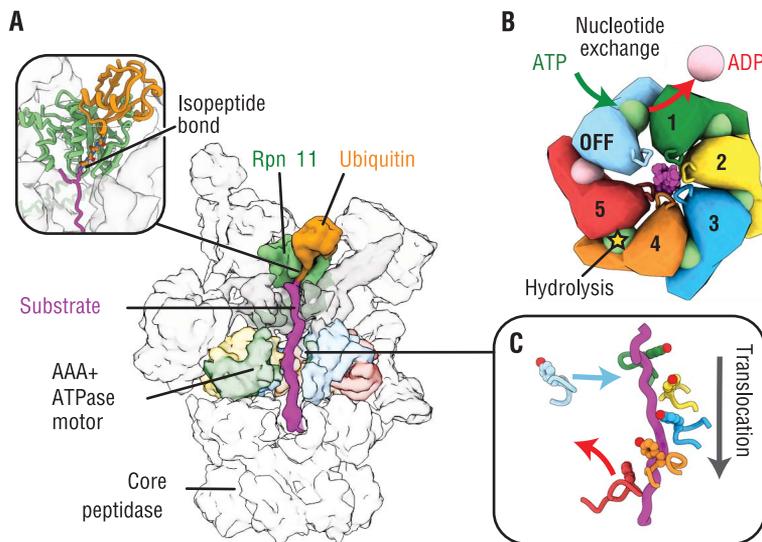
ON OUR WEBSITE

Read the full article at <http://dx.doi.org/10.1126/science.aav0725>

through the AAA+ motor, and into the core peptidase. The proteasomal motor thereby adopts staircase arrangements with five substrate-engaged subunits and one disengaged subunit. Four of the substrate-engaged subunits are ATP bound, whereas the subunit at the bottom of the staircase and the disengaged subunit are bound to adenosine diphosphate (ADP).

CONCLUSION: Of the four distinct motor states we observed, three apparently represent sequential stages of ATP binding, hydrolysis, and substrate translocation and hence reveal the coordination of individual steps in the ATPase cycle and their mechanochemical coupling with translocation. ATP hydrolysis occurs in the fourth substrate-engaged subunit from the top, concomitantly with exchange of ADP for ATP in the disengaged subunit. The subsequent transition, which is likely triggered by phosphate release from the fourth, posthydrolysis subunit of the staircase, then involves major conformational changes of the entire ATPase hexamer. The bottom ADP-bound subunit is displaced and the previously disengaged subunit binds the substrate at the top of the staircase, while the four engaged subunits move downward as a rigid body and translocate substrate toward the peptidase. Our likely consecutive proteasome conformations, together with previously determined substrate-free structures, suggest a sequential progression of ATPase subunits through the ATP-hydrolysis cycle. We hypothesize that, in general, hexameric AAA+ translocases function by this sequential mechanism. ■

Downloaded from <http://science.sciencemag.org/> on February 5, 2019



Cryo-EM structures of the substrate-engaged 26S proteasome. (A) Substrate path through the proteasome, with ubiquitin bound to Rpn11 (left inset) and the substrate polypeptide traversing through the AAA+ motor into the core peptidase. (B) Schematic showing coordinated ATP hydrolysis and nucleotide exchange observed between consecutive motor states. (C) Substrate translocation is driven by changes in the spiral-staircase arrangement of pore loops, as indicated by arrows.

The list of author affiliations is available in the full article online.
*These authors contributed equally to this work.
†Corresponding author. Email: glander@scripps.edu (G.C.L.); a.martin@berkeley.edu (A.M.)
Cite this article as A. H. de la Peña et al., *Science* 362, eaav0725 (2018). DOI: [10.1126/science.aav0725](https://doi.org/10.1126/science.aav0725)

RESEARCH ARTICLE

STRUCTURAL BIOLOGY

Substrate-engaged 26S proteasome structures reveal mechanisms for ATP-hydrolysis-driven translocation

Andres H. de la Peña^{1*}, Ellen A. Goodall^{2,3*}, Stephanie N. Gates^{2,3,4*}, Gabriel C. Lander^{1,†}, Andreas Martin^{2,3,4,†}

The 26S proteasome is the primary eukaryotic degradation machine and thus is critically involved in numerous cellular processes. The heterohexameric adenosine triphosphatase (ATPase) motor of the proteasome unfolds and translocates targeted protein substrates into the open gate of a proteolytic core while a proteasomal deubiquitinase concomitantly removes substrate-attached ubiquitin chains. However, the mechanisms by which ATP hydrolysis drives the conformational changes responsible for these processes have remained elusive. Here we present the cryo-electron microscopy structures of four distinct conformational states of the actively ATP-hydrolyzing, substrate-engaged 26S proteasome. These structures reveal how mechanical substrate translocation accelerates deubiquitination and how ATP-binding, -hydrolysis, and phosphate-release events are coordinated within the AAA+ (ATPases associated with diverse cellular activities) motor to induce conformational changes and propel the substrate through the central pore.

The 26S proteasome, the final component of the ubiquitin-proteasome system, is central to general proteostasis and the regulation of essential processes in eukaryotic cells (1). Proteins are targeted for proteasomal degradation through the covalent attachment of polyubiquitin chains to lysine residues (2). To safeguard against indiscriminate degradation, the proteolytic active sites of the proteasome are sequestered within the barrel-shaped 20S core particle (CP). Access to these active sites is controlled by the 19S regulatory particle (RP), which binds to one or both ends of the CP, recruits ubiquitinated proteins, and catalyzes their deubiquitination, unfolding, and translocation through a central pore into the proteolytic chamber of the CP for degradation (3). The RP can be further subdivided into the base and lid subcomplexes. The nine-subunit lid subcomplex fulfills important scaffolding functions and contains the Zn²⁺-dependent deubiquitinase RpnII, which is positioned above the central pore of the proteasome to remove ubiquitin chains from substrates before degradation (4–8). The base subcomplex

consists of 10 subunits, including three ubiquitin receptors and six distinct AAA+ ATPases (ATPases associated with diverse cellular activities), Rpt1 to Rpt6 (3, 9). These ATPases (adenosine triphosphatases) form a heterohexameric ring (in the order Rpt1, Rpt2, Rpt6, Rpt3, Rpt4, and Rpt5) that is the molecular motor of the proteasome (10). Each Rpt consists of a N-terminal helix, an oligonucleotide binding (OB)-fold domain, and a C-terminal AAA+ motor domain. In the heterohexamer, the N-terminal helices of neighboring Rpt pairs form a coiled coil, and the six OB-fold domains assemble into a rigid N-ring above the AAA+ motor ring (6, 8). After ubiquitin-mediated substrate recruitment, the ATPase motor engages a flexible initiation region of the substrate for subsequent mechanical translocation and unfolding (11). To facilitate substrate transfer to the CP, the ATPase hexamer also triggers opening of the CP access gate by docking conserved C-terminal tails of Rpt subunits into pockets at the surface of the CP α ring (12–14).

Like other AAA+ ATPases, the Rpt subunits contain a highly conserved nucleotide binding pocket that couples ATP binding and hydrolysis with conformational changes to produce mechanical work (15, 16). This pocket is largely formed by the signature Walker-A and Walker-B motifs, responsible for nucleotide binding and hydrolysis, respectively, and an arginine finger provided by the clockwise-neighboring ATPase subunit that coordinates the γ phosphate of ATP during hydrolysis and enables subunit communication (17). Conserved pore-1 loops protrude from each ATPase subunit into the central channel, where

they sterically interact with the substrate polypeptide and transduce nucleotide-dependent conformational changes into directional translocation (18–21).

The common functional architecture of ring-shaped hexameric helicases and AAA+ translocases gave rise to a “hand-over-hand” model for substrate translocation (22–24), which is supported by numerous cryo-electron microscopy (cryo-EM) structures of substrate-bound homohexameric AAA+ motors (25–30). These prior structures were trapped using hydrolysis-inactivating Walker-B mutations, nonhydrolyzable ATP analogs, or analogs that are slowly hydrolyzed, to reveal series of subunits in the hexamer that resemble the ATP-bound, adenosine diphosphate (ADP)-bound, and nucleotide-free states. Generally, five nucleotide-bound subunits contact the substrate polypeptide in a spiral-staircase arrangement of pore loops, whereas one subunit remains disengaged and nucleotide-free. The hand-over-hand model stems from inferences regarding how individual subunits may progress through the various nucleotide states and substrate-binding conformations around the ring. The heterohexameric proteasomal AAA+ motor adopts distinct spiral-staircase arrangements with individual Rpts in different vertical positions (6, 31–35) and thus promises more detailed insights into the progression of states during the ATP-hydrolysis and substrate-translocation cycles. However, high-resolution structural studies of the proteasome during active substrate translocation have so far been unsuccessful.

In the absence of substrate, the ATP-hydrolyzing proteasome primarily adopts the s1 state (6, 36) in which the ATPase domains of Rpt1 to Rpt6 form a spiral staircase that is not coaxially aligned with the CP and RpnII is positioned offset from the central pore of the motor. A low-resolution structure of the proteasome trapped with a stalled protein in the central pore revealed that upon substrate engagement the RP transitions from the s1 state to a processing conformation, which is characterized by a more planar ATPase ring, a rotated lid subcomplex, and a coaxial alignment of RpnII, the Rpt hexamer, and the CP (37). However, the limited resolution and strong heterogeneity of the ATPases within these stalled proteasome complexes prevented the visualization of substrate and the identification of distinct motor states.

States that share structural similarities with the substrate-processing conformation are also observed for the substrate-free proteasome as a small subpopulation in the presence of ATP (s2 state) and upon ATPase inhibition using either ATP analogs or Walker-B mutations in individual Rpt subunits (s3, s4, s5, and s6 states and the unnamed state seen in ADP-AIF_x) (14, 33–35). Cryo-EM reconstructions of these states revealed distinct spiral-staircase arrangements and nucleotide occupancies of Rpt subunits, but the lack of ATP hydrolysis and the absence of substrate limited the conclusions that could be drawn regarding the mechanisms for ATP-hydrolysis-coupled translocation.

¹Department of Integrative Structural and Computational Biology, The Scripps Research Institute, 10550 N. Torrey Pines Road, La Jolla, CA 92037, USA. ²Department of Molecular and Cell Biology, University of California, Berkeley, CA 94720, USA. ³California Institute for Quantitative Biosciences, University of California at Berkeley, Berkeley, CA 94720, USA. ⁴Howard Hughes Medical Institute, University of California at Berkeley, Berkeley, CA 94720, USA.

*These authors contributed equally to this work.

†Corresponding author. Email: glander@scripps.edu (G.C.L.); a.martin@berkeley.edu (A.M.)

We explored the mechanistic details of ATP-hydrolysis-driven substrate translocation by determining the structure of the substrate-engaged 26S proteasome in the presence of ATP. Unlike previous studies that used ATPase inhibition to trap substrate-bound states of other AAA+ motors, we stalled substrate translocation in the actively hydrolyzing motor of the proteasome by inhibiting Rpn11-mediated deubiquitination. We describe four cryo-EM structures, depicting four distinct motor states with the unambiguous assignment of substrate polypeptide traversing the RP from the lysine-attached ubiquitin at the Rpn11 active site, through the Rpt hexamer, to the gate of the CP. Three of these states appear to represent sequential stages of ATP binding, hydrolysis, and substrate translocation and hence reveal the coordination of individual steps in the ATPase cycle of the AAA+ hexamer and their mechanochemical coupling with translocation.

Four substrate-bound 26S proteasome structures

To stall translocation at a defined substrate position, we inactivated the Rpn11 deubiquitinase of *Saccharomyces cerevisiae* 26S proteasomes by incubation with the inhibitor ortho-phenanthroline (4) and added a globular substrate with a single polyubiquitinated lysine flanking an unstructured C-terminal initiation region. Proteasomes engaged the flexible initiation region and translocated the substrate until the attached ubiquitin chain reached the inhibited Rpn11, preventing further translocation and trapping the substrate in the central pore, which is indicated by a complete inhibition of degradation (fig. S1A). Stalling sub-

strate translocation in the proteasome does not also stall the AAA+ motor, as we observed a rate of ATP hydrolysis that was even slightly elevated compared with that of freely translocating proteasomes (fig. S1B). We posit that this stalled state resembles the scenario when the proteasome encounters thermodynamically stable substrate domains that require repeated pulling by the ATPase to be unfolded (31, 37).

After incubation with substrate, proteasomes were vitrified for cryo-EM single-particle analysis, which produced reconstructions of the 26S proteasome in six distinct conformational states. In the initial 3D classification, roughly 42% of particles were observed to be substrate free, adopting an s1-like state (fig. S2A and table S1), whereas the rest of the particles were sorted into reconstructions that showed ubiquitin density adjacent to Rpn11 and adopted non-s1-like conformations (fig. S2A). Further focused classification of the ATPase motor resulted in four s4-like reconstructions and one reconstruction that resembled the s2 state but lacked density for substrate within the central channel of the AAA+ motor. In contrast, the four s4-like reconstructions (ranging from ~4.2 to ~4.7 Å in overall resolution) showed clearly visible substrate density threaded through the center of the RP (Fig. 1, figs. S2 and S3A, and table S1).

Proteasome interactions with the translocating substrate

The stalled proteasome states not only revealed the detailed path of the substrate polypeptide from the RP to the CP but also resolved the structure of ubiquitin-bound Rpn11 in the context of

the 26S holoenzyme (Fig. 1A and movie S1). The most proximal, substrate-attached ubiquitin moiety of the polyubiquitin chain is positioned in the catalytic groove of Rpn11, whose ubiquitin-interacting Insert-1 region adopts the same active β -hairpin conformation previously observed in the crystal structure of the isolated ubiquitin-bound Rpn11-Rpn8 dimer (Fig. 1B and fig. S3B). Although the catalytic Zn^{2+} ion is not visible in the Rpn11 active site, likely due to the treatment with ortho-phenanthroline, the conformations of ubiquitin and Rpn11 match the active, Zn^{2+} -containing structure (Fig. S3B), with the addition of an intact isopeptide bond to the substrate lysine.

Upstream (N-terminal) of the ubiquitin-modified lysine, only two amino acids of the substrate were resolved. The orientation of these residues delineates a path near the N-terminal helix of Rpt2 by which substrates may approach the central pore of the proteasome (Fig. 1A), yet to what extent this path outside the N-ring is fixed or substrate dependent remains unclear. Downstream (C-terminal) of the ubiquitinated lysine, the substrate is confined to the narrow central channel of the Rpt hexamer (Fig. 1A and fig. S3, C and D). An axial view of the RP reveals that the Rpn11 catalytic groove is aligned with the trajectory of substrate translocation through this channel, which follows a straight line from the isopeptide bond into the AAA+ motor (fig. S3D). This alignment explains how vectorial tugging by the motor can pull ubiquitin directly into the cup-shaped Rpn11 binding site and thus accelerate cotranslocational deubiquitination (38). The active β -hairpin conformation of Rpn11's Insert-1

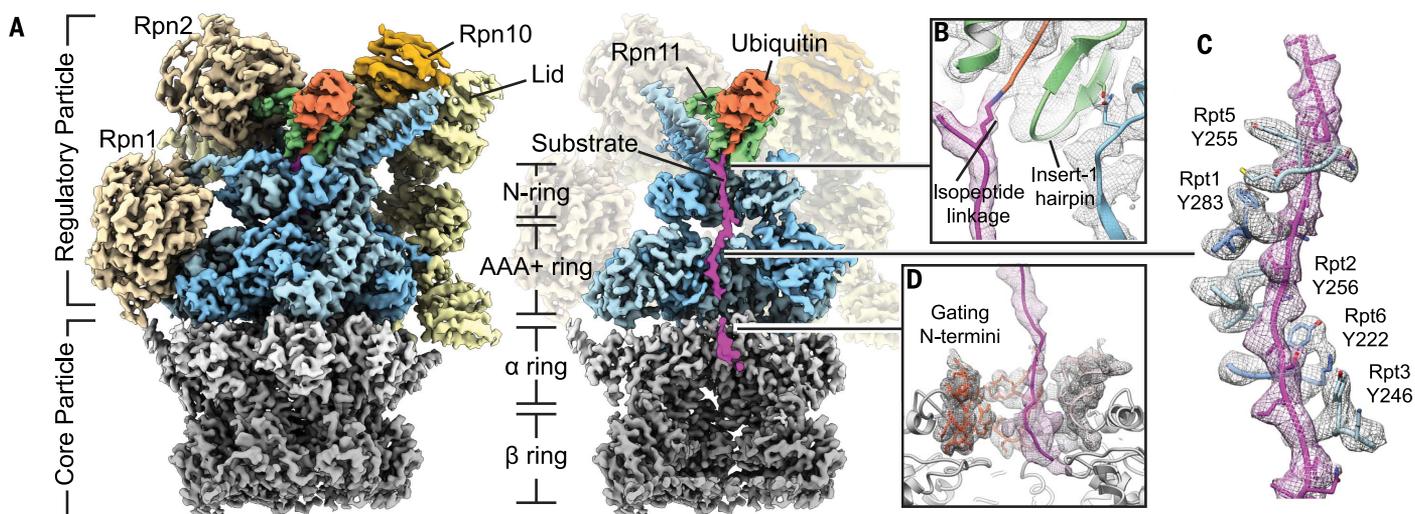


Fig. 1. High-resolution structure of the substrate-engaged 26S proteasome. (A) Exterior (left) and cutaway (right) views of the substrate-engaged proteasome cryo-EM reconstruction. The substrate (magenta) is shown extending from the ubiquitin moiety (orange), through the central pore formed by the N-ring and the AAA+ motor (blue), into the gate of the 20S core particle (gray). (B) The isopeptide bond between the substrate lysine and the C terminus of the ubiquitin moiety is bound in the catalytic

groove of Rpn11 (green), with the Insert-1 region in its active, β -hairpin state that is stabilized by a contact to the N-terminal helix of Rpt5 (blue). (C) The substrate polypeptide is encircled by a spiral staircase of pore-1 loop tyrosines (Y) projecting from the Rpt subunits. (D) Substrate enters the open gate of the core particle. The gating N termini of α subunits 2, 3, and 4 (red) extend toward the AAA+ motor, forming a hydrophobic collar in conjunction with the N termini of the other four α subunits (pink).

region appears to be stabilized through additional contacts with Rpt5 at the base of the Rpt4-Rpt5 coiled coil (Fig. 1B). Our structures suggest that the translocation stall originates from ubiquitin becoming trapped as it is pulled into the catalytic groove of inactive Rpn11, rather than sterically clashing with the narrow entrance of the N-ring. Despite having the ATPase ring in distinct hydrolysis states (see below), all four proteasome structures show ubiquitin functionally bound to Rpn11, indicating that deubiquitination indeed occurs cotranslocationally, after the regulatory particle has switched to an engaged conformation and while the substrate is threaded into the pore. This observation is consistent with a mechanism

in which the regulatory particle does not adopt a specific conformation for deubiquitination but cleaves off ubiquitin modifications as they approach Rpn11 during processive substrate translocation.

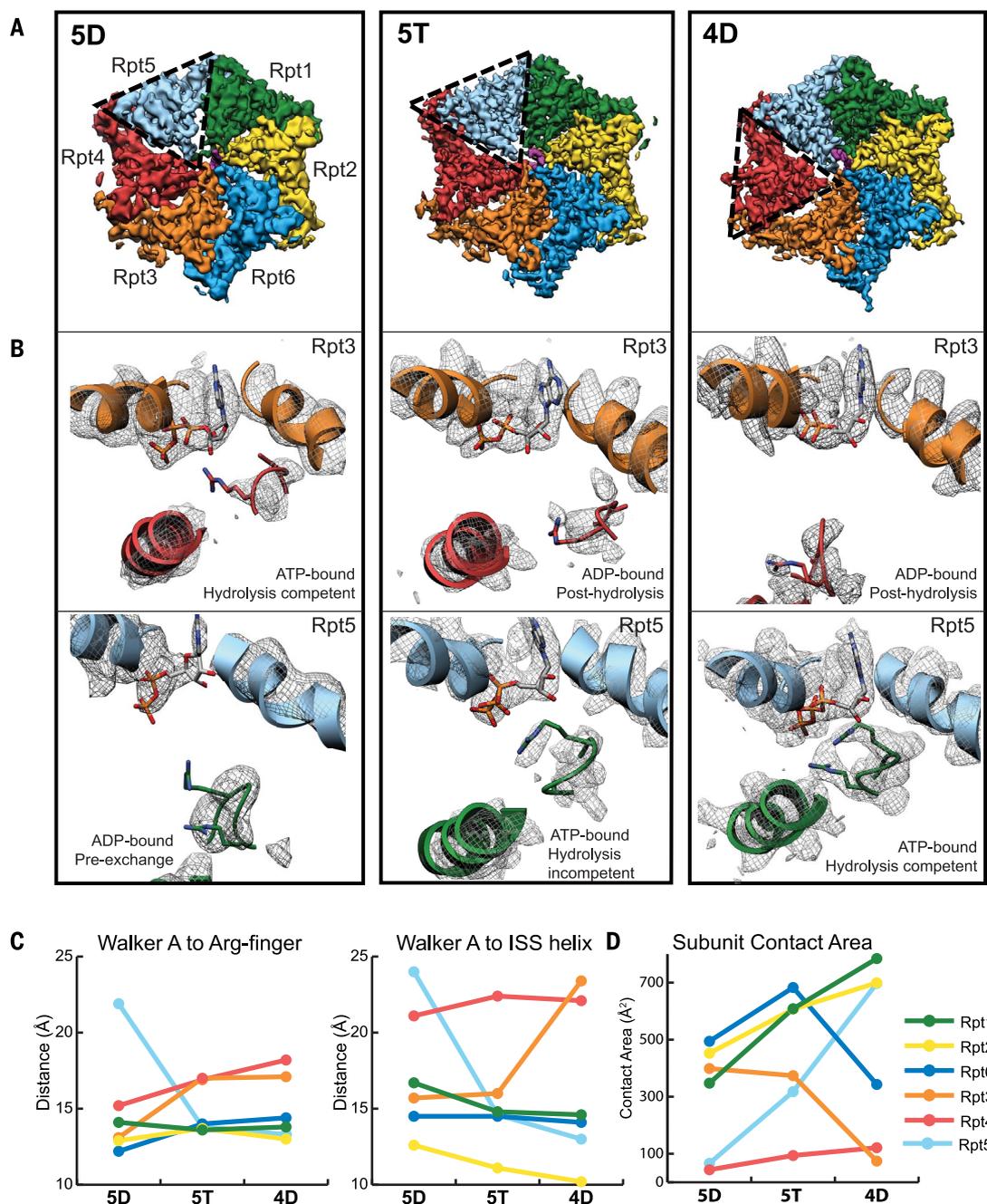
Because of the defined stall at the single ubiquitin chain, we were able to reliably model the C-terminally inserted substrate and assign a specific sequence to the polypeptide density within the AAA+ motor (Fig. 1C). The pore-1 loop Tyr and neighboring Lys residue of individual Rpt subunits form a spiral staircase that tightly encircles the substrate, consistent with a translocation mechanism that involves steric interactions with amino acid side chains of the polypeptide

(Fig. 1C). As in many other AAA+ motors, the pore-2 loops (a second loop that protrudes from each ATPase subunit into the pore) are arranged in a second staircase that lies in close proximity to the substrate below the pore-1 loop spiral (fig. S3E). In contrast to the pore-1 loops, the pore-2 loops do not contain bulky residues and may contribute to translocation through interactions of their backbones with the substrate (fig. S3E), as suggested by defects previously observed for pore-2 loop mutations (20).

After traversing the AAA+ motor, the substrate enters the gate of the CP (Fig. 1D). Our four cryo-EM structures reveal two gating conformations with distinct RP-CP interactions and

Fig. 2. Nucleotide-pocket analysis of three sequential substrate-engaged AAA+ motor conformations.

(A) Top view of AAA+ motor density maps for three sequential states, with the substrate-disengaged Rpt subunits indicated by a dashed triangle. Substrate density (magenta) is shown in the central pore formed by Rpt1 (green), Rpt2 (yellow), Rpt6 (blue), Rpt3 (orange), Rpt4 (red), and Rpt5 (light blue). (B) Close-up views of the Rpt3 nucleotide-binding pocket, showing the neighboring Rpt4 providing the Arg finger (top row) and the Rpt5 binding pocket with the Arg finger from the neighboring Rpt1 (bottom row). Individual states, left to right, are arranged in the order of motor progression. (C) Measurements of nucleotide-pocket openness colored by nucleotide-bound Rpt subunit. Shown are the distances between the α carbon of Walker-A Thr and the α carbon of the neighboring subunit's Arg finger (left) or the centroid of α -helix 10 flanking the ISS motif (right). (D) Contact area between the large AAA+ domains of neighboring Rpt subunits.



arrangements for the N termini of CP α subunits (fig. S4). In all structures, the C termini of HbYX (hydrophobic-Tyr-any amino acid)-motif containing Rpt subunits (Rpt2, Rpt3, and Rpt5) occupy the intersubunit pockets of the CP α ring, whereas the pockets for the C termini of Rpt1 and Rpt6 vary in occupancy (fig. S4A). Two of the structures show all Rpt tails except for Rpt4 docked into the intersubunit pockets and, consequently, a completely open gate, similar to previously described states in substrate-free proteasomes (14, 35) (fig. S4, B to D). In this open conformation, the gating N termini of α subunits 2, 3, and 4 become directed toward the base sub-complex and interact with the N termini of the other four α subunits through a conserved Tyr residue (fig. S4E). This results in the formation of a hydrophobic collar directly beneath the exit from the AAA+ motor (fig. S4E). The other two structures, which exhibit lower levels of Rpt1- and Rpt6-tail occupancies in the respective α -ring pockets, reveal a partially open gate (fig. S4, C and D). This observation supports a recently proposed model in which cooperative gate opening is driven by the tail insertion of Rpt1 and Rpt6, after the three HbYX-containing tails are docked (14, 35).

Distinct nucleotide states give rise to four ATPase conformations

Our four substrate-engaged proteasome structures show distinct motor conformations with nucleotide density present in all six ATP-binding pockets and one or two subunits that do not interact with substrate (Fig. 2A and fig. S5A). To reliably assign nucleotide identities and thereby establish the progression of the ATP-hydrolysis cycle within the actively hydrolyzing Rpt hexamer, we assessed not only the occupying nucleotide densities but also the geometries of the ATPase sites, the structural stability of allosteric motifs, and the intersubunit contact areas (Fig. 2 and table S2). ATP-bound, hydrolysis-competent subunits form a closed pocket with an increased intersubunit contact area characterized by a direct interaction between the γ phosphate of ATP and the well-resolved Arg fingers of the clockwise neighboring subunit (Fig. 2, B to D, and fig. S5, B to F). In contrast, ADP-bound subunits are more open with a decreased intersubunit contact area and Arg fingers that are more flexible, as indicated by lower resolvability (Fig. 2, B to D, and fig. S5, B to F). Subunits that are ATP bound but not yet hydrolysis competent and subunits where ATP hydrolysis has just occurred show similar, intermediate Arg-finger distances (Fig. 2, B and C). To distinguish between these pre- and posthydrolysis states, we assessed the pocket openness by measuring the intersubunit contact area or the distance between the conserved Walker-A motif Thr and the helix preceding the intersubunit signaling (ISS) motif of the neighboring subunit (Fig. 2, C and D, and fig. S5, B to E) (26, 33). Our analyses revealed a continuum of nucleotide states within the Rpt hexamers, ranging from ATP-bound partially open pockets with semi-engaged Arg fin-

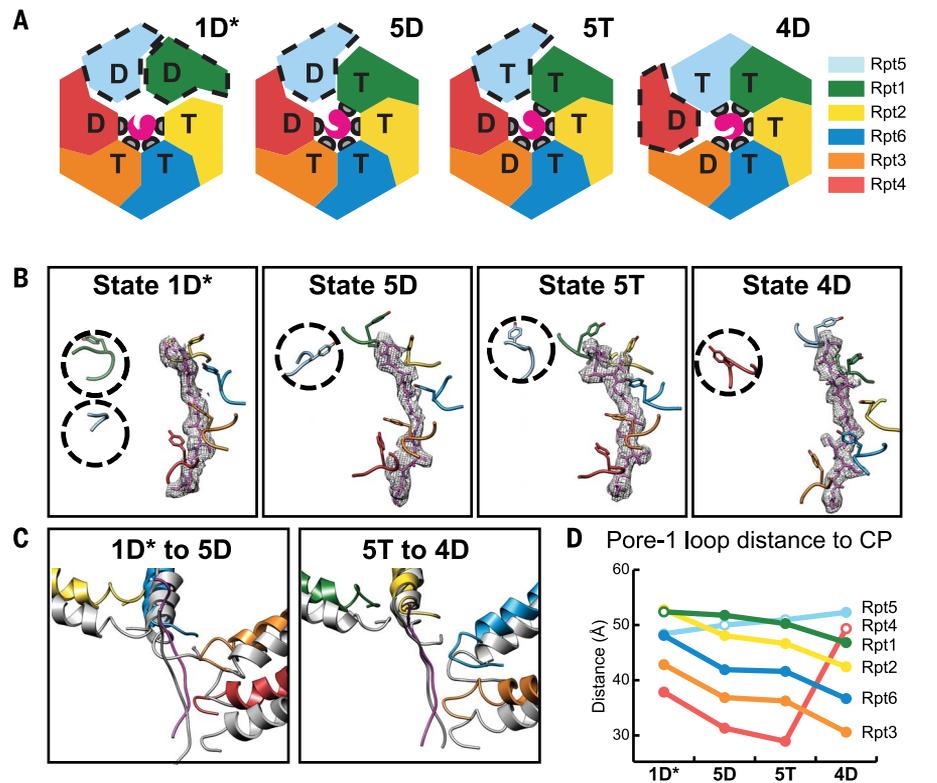


Fig. 3. Pore-1 loop tyrosines define three distinct spiral-staircase conformations of the AAA+ motor. (A) Summary of nucleotide states and staircase arrangement in the 1D*, 5D, 5T, and 4D states. Coloring of the motor subunits—Rpt1 (green), Rpt2 (yellow), Rpt6 (blue), Rpt3 (orange), Rpt4 (red), and Rpt5 (light blue)—is consistent throughout the figure. Pore-1 loop contacts (gray) with substrate (magenta) are not present in the disengaged subunits (dashed outline). (B) Pore-1 loop Tyr staircases for each of the substrate-bound states. Substrate polypeptide (mesh) is encircled by four or five engaged pore-1 loops in each state. Disengaged pore-1 loops are indicated by dashed circles. (C) Vertical movement of substrate-engaged pore-1 loops is observed during motor transition from the 1D* to 5D and 5T to 4D states. The lower state (5D and 4D, respectively) is shown in gray. (D) Plot of distances between the α carbon of the pore-1 loop tyrosines to the plane of the core particle's gate. Filled circles represent substrate-engaged pore loops; open circles indicate disengaged pore loops.

gers (hydrolysis incompetent) to ATP-bound closed pockets with fully engaged Arg fingers (hydrolysis competent), ADP-bound closed pockets with disengaged Arg fingers (posthydrolysis), and ADP-bound open pockets with disengaged Arg fingers (pre-nucleotide exchange) (Fig. 2B). Individual Rpt subunits show a progression through discrete nucleotide states around the hexameric ring (Fig. 3A), indicating that each reconstruction reflects a distinct snapshot of the proteasomal AAA+ motor during the ATPase cycle and that Rpts likely progress sequentially through this cycle. Our structures allow us to correlate the distinct vertical registers and nucleotide states of Rpt subunits and analyze the coupling of individual ATPase steps with each other and with the mechanical translocation of substrate.

Sequential motor states reveal the mechanism for ATP-hydrolysis-driven substrate translocation

The current hand-over-hand translocation model for hexameric AAA+ ATPases is based on observations of the subunits encircling and inter-

acting with substrate in a staircase-like organization, with the exception of the sixth subunit (often referred to as the “seam subunit”) that is displaced from the substrate and positioned between the lowest and highest subunits of the staircase (25–30). Consistent with this previously observed configuration, we see that in all substrate-engaged proteasome states the Rpt subunits interact with the substrate through their pore-1 loops in a spiral-staircase arrangement, with the pore-2 loops forming a similar staircase underneath (figs. S6, A and B). A characteristic seam is observed along the interface between the highest substrate-engaged subunit of the staircase and the neighboring substrate-disengaged subunit (Figs. 2A and 3, A and B, and fig. S5A). We name our four proteasome states based on the identity and the nucleotide state of this substrate-disengaged “seam” subunit: Rpt1-ADP, Rpt5-ADP, Rpt5-ATP, and Rpt4-ADP are named as 1D*, 5D, 5T, and 4D, respectively (Fig. 3A).

In these four conformations, three different Rpt subunits occupy the uppermost substrate-bound position of the staircase. In 1D*, Rpt2 is

in the top substrate-bound position with the seam subunit Rpt1 displaced from the substrate (Fig. 3B). Unexpectedly, Rpt5 is also disengaged in this conformation, indicating that 1D* may represent an off-pathway ATPase configuration (see discussion below).

On the basis of their nucleotide occupancies and spiral-staircase arrangements, we posit that the remaining three conformations—5D, 5T, and 4D—represent consecutive states whose transitions include a nucleotide exchange, a hydrolysis event, and a translocation step (Fig. 3 and movie S2). In 5D and 5T, the vertical staircase register is shifted by one subunit in the counterclockwise direction compared with 1D*, such that Rpt1 assumes the uppermost substrate-bound position, whereas the other subunits move downward and only Rpt5 is substrate disengaged (Fig. 3C). During the 5D-to-5T transition, the staircase arrangement of Rpt subunits remains largely the same (fig. S6D), but the density for the Rpt5-bound nucleotide changes concomitantly with a substantial closure of the binding pocket that brings the Arg fingers of Rpt1 into close proximity (Fig. 2B), which is consistent with an exchange of ADP for ATP. This exchange and the resulting shift of Rpt5 toward the central pore likely primes this subunit by allosterically positioning the pore loops for substrate engagement in the subsequent 4D state (Fig. 3B). Nucleotide exchange in the disengaged Rpt thus appears prerequisite for substrate binding at the top of the spiral staircase, which agrees with the highest substrate-contacting subunit always being bound to ATP (Fig. 3A). Notably, concurrent with ATP binding to Rpt5, Rpt3 hydrolyzes ATP during the 5D-to-5T transition, as indicated by correlative changes in the Rpt3-bound nucleotide density and the disengagement of the neighboring Arg finger (Fig. 2, B and C). Neither the nucleotide exchange nor the hydrolysis event cause substantial conformational changes in the Rpt hexamer, but they represent the trigger for the most pronounced rearrangement of the mechanochemical cycle in the subsequent transition to 4D.

During this 5T-to-4D transition, we observe an opening of the Rpt3 nucleotide-binding pocket and a disruption of the intersubunit interactions with the neighboring Rpt4 (Fig. 2, B and D). Rpt4 separates from Rpt3, disengages from substrate, and moves from the bottom of the staircase out and upward, which is likely driven by the topologically closed ring architecture of the Rpt hexamer. At the same time, the ATP-bound Rpt5 at the top of the staircase moves to a more central position and binds substrate (Fig. 3B), whereas the substrate-engaged subunits Rpt1, Rpt2, Rpt6, and Rpt3 move as a rigid body downward by one register and translocate the substrate toward the CP gate (Fig. 3, C and D, and movie S2).

Even though we do not detect concrete nucleotide-density changes between the 5T and 4D conformations, we can postulate based on the preceding ATP-hydrolysis event in Rpt3 and the subsequent opening of its pocket that phosphate release from Rpt3 is responsible for the disruption of intersubunit interactions with Rpt4

and the consequent conformational changes of the entire ATPase ring. This model is consistent with our observations that the penultimate subunit in the staircase exhibits a completely or partially closed pocket in all proteasome conformations, whereas the lowest substrate-engaged subunit is always ADP bound with an open pocket (Figs. 3A; and 2, C and D; and fig. S5, B to E). Furthermore, it agrees with previous single-molecule data on the homohexameric ClpX ATPase, suggesting that phosphate release represents the force-generating step of the ATPase cycle (39). Similar to the coordinated nucleotide-exchange and ATP-hydrolysis steps in the previous 5D-to-5T transition, the disruption of the Rpt3-Rpt4 interface through potential phosphate release, the substrate-engagement by Rpt5, and the movement of a four-subunit rigid body for substrate translocation appear to be interdependent and tightly coupled during the 5T-to-4D transition.

For the ATP-hydrolysis and substrate-translocation cycles, our findings suggest that a particular subunit binds ATP and engages substrate at the uppermost position, hydrolyzes ATP when at the penultimate position of the staircase, releases phosphate as it moves to the bottom of the ring, and disengages from substrate in the next step (Fig. 4A). AAA+ motor movements and substrate translocation would thus be powered by sequential ATP hydrolysis and phosphate release as each Rpt transitions to the bottom of the staircase. Our observation of four substrate-engaged subunits moving as a rigid body to translocate substrate in response to ATP hydrolysis and phosphate release is consistent with previous biochemical studies of the ClpX ATPase, which indicated that several subunits interact synergistically with substrate, allowing even a pore-1 loop-deficient subunit to drive translocation (40, 41). The rigid-body movement of four Rpts vertically advances the engaged pore-1 loop Tyr residues by ~6 Å (Fig. 3C and D), suggesting a fundamental step size of two amino acids per hydrolyzed ATP for proteasomal substrate translocation.

We do not observe a vertical movement of substrate due to the defined stall of translocation upon Rpn11 inhibition, and all of our proteasome conformations show largely the same stretch of polypeptide in the central channel. Nevertheless, the substrate responds to staircase rearrangements with lateral movements in the ATPase channel, shifting toward the engaged pore-1 loops and away from the disengaged subunits (fig. S6A). The substrate backbone follows the spiral-staircase arrangement of pore loops rather than traversing the motor in a straight vertical path, and its lateral position in the channel rotates counterclockwise around the hexamer as the Rpts progress through the various nucleotide states (fig. S6A).

Additional states of the proteasomal ATPase cycle

Whereas 5D, 5T, and 4D each contain four ATP-bound and two ADP-bound subunits, 1D* shows three ATP-bound and three ADP-bound subunits (Fig. 3A). We interpret this conformation as an

alternate, potentially off-pathway version of a 1D state. A comparison of the Rpt subunit organization in 1D* with those in the 5D, 5T, and 4D states, as well as other proteasome and AAA+ motor structures (figs. S7, A and B), suggests that Rpt5 has prematurely released from substrate at the bottom of the spiral after opening of the Rpt4 nucleotide-binding pocket, and hence both Rpt5 and Rpt1 are disengaged (Fig. 3B and fig. S7A). The Rpt5 pore-1 loop is divergent from the other Rpts, containing a conserved Met rather than a Lys (42), which could result in weaker substrate interactions, especially when translocation is stalled, and thus contribute to the premature disengagement in the 1D* state. Conversely, the 1D* state might be explained by failed nucleotide exchange in Rpt1 at the top of the spiral, which would prevent substrate engagement and the consequent rearrangement of the staircase to the 5D state. Notably, the previously described s3 conformation of the substrate-free proteasome shows the expected staircase arrangement of 1D, with Rpt5 remaining in the lowest position of the staircase (fig. S7B) (33).

Our three distinct spiral-staircase states offer a view of the discrete events leading to a complete step of hydrolysis-driven substrate translocation, yet the current hand-over-hand model requires that every Rpt subunit cycles through all the states. Previous biochemical studies have indicated that ATP hydrolysis in almost all Rpt subunits contribute to substrate engagement and translocation (20, 43, 44). Therefore, the proteasome conformations described here likely represent only a subset of states that may be complemented by the corresponding 4T, 3D, 3T, 6D, 6T, 2D, and 2T states, as well as an additional 1T state between 1D and 5D (Fig. 4, B to D, and fig. S7), to complete the ATP-hydrolysis and substrate-translocation cycles of the proteasome.

Indeed, several additional staircases have been previously observed for the proteasome, albeit in the absence of substrate and induced by non-hydrolyzable ATP analogs, which hampered robust conclusions about mechanochemical coupling or the ATPase cycle. Their overall similarity to our Rpt staircase arrangements is sufficient to designate specific spiral-staircase states. Some of these states (e.g., the ADP- AlF_4^- -bound and s2 states) were regarded as unlikely processing conformations of the AAA+ motor, as they were associated with a partially open CP gate (14, 33, 34). However, two of our engaged states similarly contain only partially open gates yet clearly show substrate being threaded through the central channel to the CP gate (fig. S4). This indicates that a fully open gate is not required for every step of substrate translocation, but its openness may vary depending on the state of the Rpt staircase and corresponding allosteric subtleties in Rpt-tail interactions with the CP. More predictive criteria for a processing motor state are the coaxial alignment of the AAA+ motor with CP, the rotation of the lid subcomplex, and the presence of rigid bodies formed between the large AAA subdomain of one subunit and the small AAA subdomain of its neighbor in all but

the substrate-disengaged Rpts. On the basis of these criteria and their staircase orientation, the s2 and recently described s5 states (14) would represent 3D or 3T states, and the ADP-AIF_x-bound proteasome conformation (34) resembles a putative 2D* or 2T* state, with two substrate-disengaged subunits similar to 1D* (Fig. 4, B and C). The s4 and S_{D2} states, which had previously been proposed as potential processing states (33, 35), show overall staircase similarities with our substrate-engaged 4D and 5D states, respectively, even though some of their Rpts are distorted as a likely consequence of inhibited ATP hydrolysis and the absence of substrate (fig. S7, C and D).

Our substrate-engaged proteasome states provide a structural context for previously described ATP analog-bound conformations, enabling us to approximate all possible Rpt staircases, except for the 6D and 6T states, and thus support the model of a sequential hand-over-hand mechanism wherein each Rpt transitions through the ATP-hydrolysis and substrate-translocation cycles (Fig. 4D). Why the proteasome in the presence of substrate preferentially adopts only the 1D*, 5D, 5T, and 4D states remains unclear; however, the likely consecutive 5D, 5T, and 4D states are sufficient to provide us with a complete picture of subunit transitions during the ATPase cycle and substrate translocation.

Outlook

We elucidated structures of the substrate-engaged 26S proteasome that answer many of the outstanding questions regarding proteasomal degradation and the general mechanism by which AAA+ translocases process their substrates. Inhibiting deubiquitination by Rpn11 led to a trapped state in which the substrate-attached ubiquitin remains functionally bound in the Rpn11 catalytic groove and the scissile isopeptide bond is linearly aligned with the translocation trajectory through the AAA+ motor (Fig. 1). We conclude that during normal degradation, ubiquitin modifications are pulled directly into the Rpn11 catalytic groove. This ubiquitin-capture mechanism explains how Rpn11 functions as a gatekeeper to efficiently remove all ubiquitin modifications from a substrate during processive translocation, as well as how deubiquitination can be accelerated by mechanical pulling of the AAA+ motor on the substrate polypeptide (38).

We resolved three apparently sequential states of the Rpt heterohexamer, which provide a model for the intersubunit coordination during nucleotide exchange, ATP hydrolysis, and phosphate release within the AAA+ motor, as well as how these events are mechanochemically coupled to substrate translocation. Consistent with other substrate-bound AAA+ ATPase structures, the proteasome motor adopts staircase arrangements that encircle the unfolded polypeptide substrate, with one subunit disengaged (25–29, 45). Four of the substrate-engaged subunits are ATP bound, whereas the subunit at the bottom of the staircase and the disengaged subunit are bound to ADP. Our structures suggest that nucleotide ex-

change primes the disengaged subunit for substrate binding at the top of the staircase and that this exchange occurs concomitantly with ATP hydrolysis in the fourth substrate-engaged subunit from the top. Both steps of the ATPase cycle are associated with only subtle allosteric rearrangements, whereas the entire ATPase hexamer undergoes major conformational changes during the subsequent transition that appears to be linked to phosphate release from the posthydrolysis, fourth subunit of the staircase. These rear-

rangements include the displacement of the bottom ADP-bound subunit, substrate binding of the previously disengaged subunit at the top of the staircase, and the downward movement of the remaining four substrate-engaged subunits as a rigid body. It appears that all of these processes must happen in a coordinated fashion for substrate translocation to occur.

The likely consecutive ATPase states that we observe, together with equivalent staircase arrangements in previous substrate-free proteasome

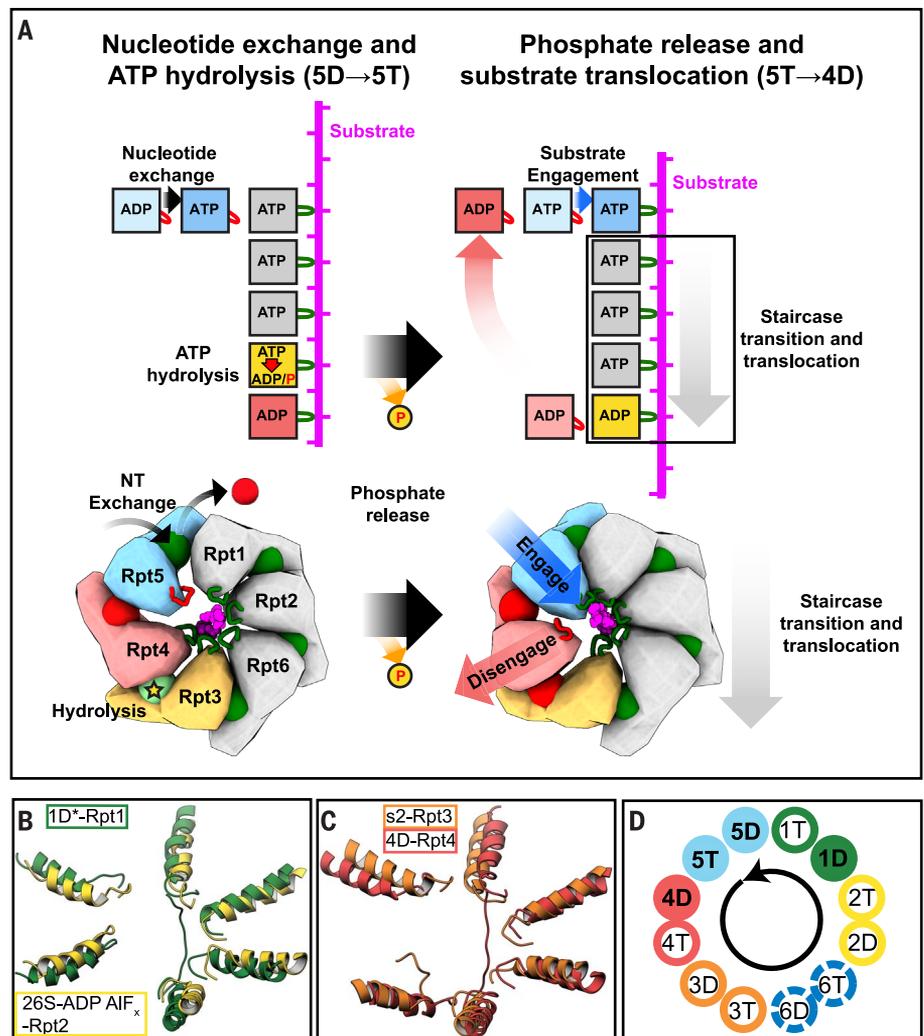


Fig. 4. Coordinated ATP-hydrolysis and substrate-translocation cycles of the proteasome.

(A) Model for the coordination of ATP-hydrolysis steps and their coupling to substrate translocation. Nucleotide exchange and ATP hydrolysis occur simultaneously in the substrate-disengaged (blue) and penultimate subunit (orange) of the staircase, respectively, with no major conformational changes of the motor. Subsequent phosphate release from the penultimate subunit leads to the displacement of the bottom subunit (red), substrate-engagement by the top subunit (blue), and downward movement of a four-subunit rigid body (boxed) to translocate substrate. (B) Spiral staircases of 1D* (dark green) and the 26S-ADP-AIF_x [yellow; PDB:5wvk (35)], rotated to overlay the disengaged subunits (1D*-Rpt1 and 26S-ADP-AIF_x-Rpt2) and aligned by the pore-1 loop helices. (C) Spiral staircases of the s2 state [orange; PDB: (33)] and the 4D state, rotated to overlay the disengaged subunits (s2-Rpt3 and 4D-Rpt4) and aligned by the pore-1 loop helices. (D) Schematic of the possible progression of proteasome states, colored by the disengaged subunit, with our observed staircase states indicated by filled circles, staircases from substrate-free proteasome states indicated by open circles, and potential additional states represented by dashed circles.

structures, suggest a sequential progression of individual Rpt subunits through the ATPase cycle, rather than a burst mechanism, where several subunits hydrolyze in rapid succession before nucleotide exchange, as proposed for the ClpX motor on the basis of single-molecule measurements (46, 47). Given the structural and functional similarities between the proteasomal Rpt hexamer and other AAA+ motors, we hypothesize that this sequential ATP-hydrolysis and substrate-translocation mechanism applies to hexameric AAA+ translocases in general. It is reminiscent of a six-subunit conveyor belt, in which a four-subunit rigid body grips the substrate and moves downward as the bottommost subunit disengages and the topmost subunit reengages substrate (Fig. 4A). The coordinated gripping by pore loops of four subunits, which are stabilized by ATP-bound, closed interfaces, likely enables higher pulling forces and reduced slippage, consistent with previous biochemical studies of the ClpX motor (40, 41). Similar conveyor-belt mechanisms have been proposed previously for AAA+ protein translocases as well as DNA and RNA helicases (22, 23, 26–29, 45), yet our structures clarify the precise movement of ATPase subunits and their coordination with individual steps of the ATP-hydrolysis cycle.

Materials and methods

Sample preparation

Purification of proteasome holoenzyme

26S proteasomes were purified from strain YYS40 [*MATa leu2-3,112 trp1-1 can1-100 ura3-lade2-1 his3-11,15 RPN1::RPN1-3XFLAG (HIS3)*] (48) as previously described (49). Briefly, frozen yeast paste from saturated cultures was lysed in a Spex SamplePrep 6875 Freezer/Mill, and cell powder was resuspended in 60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 8 mM MgCl₂, 2.5% glycerol, 0.2% NP-40, and ATP regeneration mix (5 mM ATP, 0.03 mg/ml creatine kinase, 16 mM creatine phosphate). Proteasomes were batch-bound to anti-FLAG M2 Affinity Gel (Millipore Sigma), washed with Wash Buffer (60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 8 mM MgCl₂, 2.5% glycerol, 5 mM ATP), eluted with 3XFLAG peptide, and further separated by size-exclusion chromatography using a Superose 6 Increase column in 60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 10 mM MgCl₂, 2.5% glycerol, and 1 mM ATP.

Preparation of ubiquitinated model substrate

A model substrate consisting of an N-terminal Cys, lysine-less titin-I27^{V15P}, a single-lysine-containing sequence derived from an N-terminal fragment of *Sphaerechinus granularis* cyclinB (residues 22 to 42, with Lys-to-Ala substitutions), a Rsp5 recognition motif (PPPY), and 6X His-tag, was purified after expression in *Escherichia coli* BL21-Star by standard methods. Briefly, In Terrific Broth, protein expression was induced with IPTG at OD₆₀₀ = 1.2 to 1.5 for 5 hours at 30°C. Cells were harvested by centrifugation, resuspended in chilled lysis buffer (60 mM HEPES,

pH 7.6 100 mM NaCl, 100 mM KCl, 15 mM imidazole) and lysed by sonication. Following clarification by centrifugation at 20,000 × g, the protein was purified using Ni-NTA affinity chromatography. The substrate was fluorescently labeled using 5-fluorosecin maleimide at pH 7.2 for 3 hours at room temperature and quenched with DTT. Free dye was separated from the substrate by size-exclusion chromatography with a Superdex 200 column (GE Healthcare), buffer exchanging the substrate into 60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 10 mM MgCl₂, 2.5% glycerol.

The substrate at final substrate concentration of 50 μM was modified with long, K63-linked ubiquitin chains using 5 μM *Mus musculus* Uba1, 5 μM *S. cerevisiae* Ubc1, 20 μM *S. cerevisiae* Rsp5ΔWW (20, 50), and 2 mM *S. cerevisiae* ubiquitin, in 60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 10 mM MgCl₂, 2.5% glycerol, and 15 mM ATP for 3 hours at 25°C, followed by incubation overnight at 4°C.

ATPase assay

Proteasome ATPase activity was monitored using a spectrophotometric assay that couples regeneration of hydrolyzed ATP to the oxidation of NADH (51). Reactions contained a final concentration of 150 nM 26S proteasome that had been preincubated with ortho-phenanthroline and ATPase mix for 5 min on ice or mock treated before bringing the sample to 25°C and adding FAM-labeled ubiquitinated substrate to a final concentration of 3 μM and ortho-phenanthroline to a final concentration of 3 mM. Absorbance at 340 nm was measured for 10 min with 12-s intervals in a 384-well plate (Corning) using a Biotek Synergy Neo2 plate reader. Reactions were done in 60 mM HEPES, pH 7.6, 20 mM NaCl, 20 mM KCl, 10 mM MgCl₂, 2.5% glycerol 1 mM TCEP, and 1 X ATPase mix (5 mM ATP, 3 U ml⁻¹ pyruvate kinase, 3 U ml⁻¹ lactate dehydrogenase, 1 mM NADH, and 7.5 mM phosphoenol pyruvate).

Gel-based and fluorescence anisotropy-based monitoring of proteasome degradation

200 nM proteasome was pre-incubated with 3mM ortho-phenanthroline as described in the ATPase assay. Upon the addition of substrate, fluorescence anisotropy of the substrate-attached FAM dye was measured with a 5-s interval in a 384-well plate (Corning) using a Biotek Synergy Neo2 plate reader. 10 min after the addition of ubiquitinated substrate, samples were quenched by the addition of 2% SDS and separated on a 4 to 20% gradient Tris-Glycine SDS-PAGE gel (BioRad). Fluorescence at 530nm from the FAM-labeled substrate was measured using a BioRad ChemiDoc MP imager.

Grid preparation for cryo-electron microscopy

26S proteasomes were diluted to a concentration of 20 μM in a solution containing 20 mM HEPES, pH 7.6, 25 mM NaCl, 25 mM KCl, 10 mM

MgCl₂, 1 mM TCEP, 0.05% NP-40, an ATP regeneration mix (5 mM ATP, 0.03 mg/ml creatine kinase, 16 mM creatine phosphate), and 6 mM ortho-phenanthroline. This solution was mixed with an equal volume of 50 μM ubiquitinated model substrate. Three microliters of the holoenzyme-substrate solution were immediately applied to R2/2 400-mesh grids (Quantifoil) that had been plasma treated for 20 s using a glow discharger (Electron Microscopy Sciences) operated under atmospheric gases. The grids were manually blotted to near dryness with Whatman no. 1 filter paper inside a cold room (4°C) and gravity plunged into liquid ethane using a home-built system.

Data collection and image processing

Cryo-EM data were acquired using the Leginon software for automated data acquisition (52) and a Titan Krios (Thermo Fisher) equipped with a K2 Summit (Gatan) direct electron detector in counting mode (table S1). Movies were collected by navigating to the center of a hole and sequentially image shifting to 10 targets situated at the periphery of the 2-μm hole (fig. S2F). To maximize the number of targets per hole, a nanoprobe beam of 597 nm in diameter was utilized. This resulted in a total acquisition of 11,656 movies at an approximate rate of 2200 movies per day. Movies were recorded at a nominal magnification of 29000x (1.03-Å magnified pixel size) and composed of 25 frames (250 ms per frame, ~50 e⁻/Å² per movie). Movie collection was guided by real-time assessment of image and vitrified sample quality using the Appion image-processing software (53). Frame alignment and dose weighting were performed in real-time using UCSF Motioncor2 (54). CTF estimation on aligned, unweighted, micrographs was performed with Getf (55).

All data postprocessing steps were conducted in RELION 2.1 (56, 57). Holoenzyme particles were picked using s1 proteasome templates generated from 2D class averages obtained from a prior cryo-EM experiment. This resulted in 579,361 particle picks that were extracted (660 pixels × 660 pixels) and downsampled (110 pixels × 110 pixels) for reference-free 2D classification. 298,997 particles, belonging to the 2D classes demonstrating features characteristic of secondary structural elements, were subjected to 3D refinement and subsequent 3D classification (*k* = 10). A 3D template of an s1 proteasome was utilized to guide the initial 3D refinement and 3D classification, which ensured that s4-like or substrate-bound reconstructions did not arise from template bias. 238,828 particles corresponding to 3D classes without artefactual features were chosen for further data processing. To minimize the detrimental effects of the holoenzyme's pseudo-symmetry (C2) on resolution, the raw holoenzyme particles were C2 symmetry expanded, 3D refined, and a python script was used to determine the *x* and *y* coordinates corresponding to the center of the ATPase within the regulatory particles (RP). In this way, the RPs at each end of every core particle were re-extracted to serve as individual

asymmetric units without down-sampling. A round of reference-free 2D classification enabled us to remove the ends of core particles that lacked a regulatory particle. This combined expansion and classification approach netted 380,011 distinct RP-containing particles.

We performed 3D classification on the RP dataset and isolated 242,980 particles whose parent 3D class exhibited a globular ubiquitin-shaped density in the periphery of the Rpn11 active site. Further classification aimed to identify substrate in the central pore of the proteasome. To accomplish this, a soft mask encompassing the AAA+ motor was used to exclude the rest of the proteasome for 3D classification and 3D refinement. This resulted in four distinct AAA+ motor reconstructions containing density attributed to substrate in the central pore with nominal resolutions ranging from 3.9 to 4.7 Å (fig. S2). To further increase map quality outside the AAA+ motor, the global maps corresponding to each AAA+ motor were subdivided into 12 regions for focused 3D refinement, and a composite map consisting of all 12 focused 3D refinements was then generated for each reconstruction to facilitate atomic model building (fig. S2E).

Atomic model building

All atomic models were built using the s4 proteasome model [PDB ID: 5MPC (33)] as a template. The initial template's subunits were individually rigid body fit into each of the four EM reconstructions (1D*, 5D, 5T, and 4D) with Chimera "Fit in Map" (58). The docked templates were then subjected to one cycle of morphing and simulated annealing in PHENIX, followed by a total of 10 real-space refinement macrocycles utilizing atomic displacement parameters, secondary structure restraints (xssdssp), local grid searches, and global minimization (59). After automated PHENIX refinement, manual real-space refinement was performed in Coot (60). Residue side chains without attributable density were truncated at the α -carbon, ions were removed, and atoms corresponding to the β -ring, N-ring, and RP lid were removed in the 1D*, 5D, and 5T models due to redundancy and to accelerate refinement. For the 4D state, a similar approach was followed, but atoms corresponding to the N-ring were not removed to facilitate template-based (PDB ID: 5U4P) (38) modeling of ubiquitin at the Rpn11 active site. Isopeptide bond-length restraints were manually created and implemented during PHENIX refinement (59). Multiple rounds of real-space refinement in PHENIX (five macro cycles, no morphing, no simulated annealing) and Coot were performed to address geometric and steric discrepancies identified by the RCSB PDB validation server and MolProbity (59–61). To ensure atomic models were not overfit by simulated annealing, morphing, and real space refinement, map-model FSCs were calculated with PHENIX (fig. S2H).

All images were generated using UCSF Chimera (58) and ChimeraX (62).

REFERENCES AND NOTES

- G. A. Collins, A. L. Goldberg, The Logic of the 26S Proteasome. *Cell* **169**, 792–806 (2017). doi: [10.1016/j.cell.2017.04.023](https://doi.org/10.1016/j.cell.2017.04.023); pmid: 28525752
- D. Komander, M. Rape, The ubiquitin code. *Annu. Rev. Biochem.* **81**, 203–229 (2012). doi: [10.1146/annurev-biochem-060310-170328](https://doi.org/10.1146/annurev-biochem-060310-170328); pmid: 22524316
- J. A. M. Bard *et al.*, Structure and Function of the 26S Proteasome. *Annu. Rev. Biochem.* **87**, 697–724 (2018). doi: [10.1146/annurev-biochem-062917-011931](https://doi.org/10.1146/annurev-biochem-062917-011931); pmid: 29652515
- T. Yao, R. E. Cohen, A cryptic protease couples deubiquitination and degradation by the proteasome. *Nature* **419**, 403–407 (2002). doi: [10.1038/nature01071](https://doi.org/10.1038/nature01071); pmid: 12353037
- R. Verma *et al.*, Role of Rpn11 metalloprotease in deubiquitination and degradation by the 26S proteasome. *Science* **298**, 611–615 (2002). doi: [10.1126/science.1075898](https://doi.org/10.1126/science.1075898); pmid: 12183636
- G. C. Lander *et al.*, Complete subunit architecture of the proteasome regulatory particle. *Nature* **482**, 186–191 (2012). doi: [10.1038/nature10774](https://doi.org/10.1038/nature10774); pmid: 22372024
- M. H. Glickman *et al.*, A subcomplex of the proteasome regulatory particle required for ubiquitin-conjugate degradation and related to the COP9-signalosome and eIF3. *Cell* **94**, 615–623 (1998). doi: [10.1016/S0092-8674\(00\)81603-7](https://doi.org/10.1016/S0092-8674(00)81603-7); pmid: 9741626
- K. Lasker *et al.*, Molecular architecture of the 26S proteasome holocomplex determined by an integrative approach. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 1380–1387 (2012). doi: [10.1073/pnas.1120559109](https://doi.org/10.1073/pnas.1120559109); pmid: 22307589
- M. H. Glickman, D. M. Rubin, V. A. Fried, D. Finley, The regulatory particle of the *Saccharomyces cerevisiae* proteasome. *Mol. Cell. Biol.* **18**, 3149–3162 (1998). doi: [10.1128/MCB.18.6.3149](https://doi.org/10.1128/MCB.18.6.3149); pmid: 9584156
- R. J. Tomko Jr., M. Funakoshi, K. Schneider, J. Wang, M. Hochstrasser, Heterohexameric ring arrangement of the eukaryotic proteasomal ATPases: Implications for proteasome structure and assembly. *Mol. Cell* **38**, 393–403 (2010). doi: [10.1016/j.molcel.2010.02.035](https://doi.org/10.1016/j.molcel.2010.02.035); pmid: 20471945
- T. Inobe, S. Fishbain, S. Prakash, A. Matouschek, Defining the geometry of the two-component proteasome degron. *Nat. Chem. Biol.* **7**, 161–167 (2011). doi: [10.1038/nchembio.521](https://doi.org/10.1038/nchembio.521); pmid: 21278740
- D. M. Smith *et al.*, Docking of the proteasomal ATPases' carboxyl termini in the 20S proteasome's alpha ring opens the gate for substrate entry. *Mol. Cell* **27**, 731–744 (2007). doi: [10.1016/j.molcel.2007.06.033](https://doi.org/10.1016/j.molcel.2007.06.033); pmid: 17803938
- S. Chen *et al.*, Structural basis for dynamic regulation of the human 26S proteasome. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 12991–12996 (2016). doi: [10.1073/pnas.1614614113](https://doi.org/10.1073/pnas.1614614113); pmid: 27791164
- M. R. Eisele *et al.*, Expanded Coverage of the 26S Proteasome Conformational Landscape Reveals Mechanisms of Peptidase Gating. *Cell Reports* **24**, 1301–1315.e5 (2018). doi: [10.1016/j.celrep.2018.07.004](https://doi.org/10.1016/j.celrep.2018.07.004); pmid: 30067984
- J. P. Erzberger, J. M. Berger, Evolutionary relationships and structural mechanisms of AAA+ proteins. *Annu. Rev. Biophys. Biomol. Struct.* **35**, 93–114 (2006). doi: [10.1146/annurev.biophys.35.040405.101933](https://doi.org/10.1146/annurev.biophys.35.040405.101933); pmid: 16689629
- P. I. Hanson, S. W. Whiteheart, AAA+ proteins: Have engine, will work. *Nat. Rev. Mol. Cell Biol.* **6**, 519–529 (2005). doi: [10.1038/nrm1684](https://doi.org/10.1038/nrm1684); pmid: 16072036
- P. Wendler, S. Ciniawsky, M. Kock, S. Kube, Structure and function of the AAA+ nucleotide binding pocket. *Biochim. Biophys. Acta* **1823**, 2–14 (2012). doi: [10.1016/j.bbmr.2011.06.014](https://doi.org/10.1016/j.bbmr.2011.06.014); pmid: 21839118
- J. Hinnerwisch, W. A. Fenton, K. J. Furtak, G. W. Farr, A. L. Horwich, Loops in the central channel of ClpA chaperone mediate protein binding, unfolding, and translocation. *Cell* **121**, 1029–1041 (2005). doi: [10.1016/j.cell.2005.04.012](https://doi.org/10.1016/j.cell.2005.04.012); pmid: 15989953
- A. Martin, T. A. Baker, R. T. Sauer, Pore loops of the AAA+ ClpX machine grip substrates to drive translocation and unfolding. *Nat. Struct. Mol. Biol.* **15**, 1147–1151 (2008). doi: [10.1038/nstruc.2008.153](https://doi.org/10.1038/nstruc.2008.153); pmid: 18931677
- R. Beckwith, E. Estrin, E. J. Worden, A. Martin, Reconstitution of the 26S proteasome reveals functional asymmetries in its AAA+ unfoldase. *Nat. Struct. Mol. Biol.* **20**, 1164–1172 (2013). doi: [10.1038/nstruc.2013.025](https://doi.org/10.1038/nstruc.2013.025); pmid: 24013205
- J. Eroles, M. A. Hoyt, F. Troll, P. Coffino, Functional asymmetries of proteasome translocase pore. *J. Biol. Chem.* **287**, 18535–18543 (2012). doi: [10.1074/jbc.M112.357327](https://doi.org/10.1074/jbc.M112.357327); pmid: 22493437
- N. D. Thomsen, J. M. Berger, Running in reverse: The structural basis for translocation polarity in hexameric helicases. *Cell* **139**, 523–534 (2009). doi: [10.1016/j.cell.2009.08.043](https://doi.org/10.1016/j.cell.2009.08.043); pmid: 19879839
- E. J. Enemark, L. Joshua-Tor, Mechanism of DNA translocation in a replicative hexameric helicase. *Nature* **442**, 270–275 (2006). doi: [10.1038/nature04943](https://doi.org/10.1038/nature04943); pmid: 16855583
- A. Martin, T. A. Baker, R. T. Sauer, Rebuilt AAA+ motors reveal operating principles for ATP-fueled machines. *Nature* **437**, 1115–1120 (2005). doi: [10.1038/nature04031](https://doi.org/10.1038/nature04031); pmid: 16237435
- S. N. Gates *et al.*, Ratchet-like polypeptide translocation mechanism of the AAA+ disaggregase Hsp104. *Science* **357**, 273–279 (2017). doi: [10.1126/science.aan1052](https://doi.org/10.1126/science.aan1052); pmid: 28619716
- C. Puchades *et al.*, Structure of the mitochondrial inner membrane AAA+ protease YME1 gives insight into substrate processing. *Science* **358**, ea00464 (2017). doi: [10.1126/science.aao0464](https://doi.org/10.1126/science.aao0464); pmid: 29097521
- N. Monroe, H. Han, P. S. Shen, W. I. Sundquist, C. P. Hill, Structural basis of protein translocation by the Vps4-Vta1 AAA ATPase. *eLife* **6**, e24487 (2017). doi: [10.7554/eLife.24487](https://doi.org/10.7554/eLife.24487); pmid: 28379137
- Z. A. Ripstein, R. Huang, R. Augustyniak, L. E. Kay, J. L. Rubinstein, Structure of a AAA+ unfoldase in the process of unfolding substrate. *eLife* **6**, e25754 (2017). doi: [10.7554/eLife.25754](https://doi.org/10.7554/eLife.25754); pmid: 28390173
- C. Deville *et al.*, Structural pathway of regulated substrate transfer and threading through an Hsp100 disaggregase. *Sci. Adv.* **3**, e1701276 (2017). doi: [10.1126/sciadv.1701276](https://doi.org/10.1126/sciadv.1701276); pmid: 28798962
- C. Alfieri, L. Chang, D. Barford, Mechanism for remodelling of the cell cycle checkpoint protein MAD2 by the ATPase TRIP13. *Nature* **559**, 274–278 (2018). doi: [10.1038/s41586-018-0281-1](https://doi.org/10.1038/s41586-018-0281-1); pmid: 29973720
- M. E. Matyskiela, G. C. Lander, A. Martin, Conformational switching of the 26S proteasome enables substrate degradation. *Nat. Struct. Mol. Biol.* **20**, 781–788 (2013). doi: [10.1038/nstruc.2013.025](https://doi.org/10.1038/nstruc.2013.025); pmid: 23770819
- P. Ślędz *et al.*, Structure of the 26S proteasome with ATP-γS bound provides insights into the mechanism of nucleotide-dependent substrate translocation. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 7264–7269 (2013). doi: [10.1073/pnas.1305782110](https://doi.org/10.1073/pnas.1305782110); pmid: 23589842
- M. Wehmer *et al.*, Structural insights into the functional cycle of the ATPase module of the 26S proteasome. *Proc. Natl. Acad. Sci. U.S.A.* **114**, 1305–1310 (2017). doi: [10.1073/pnas.1621129114](https://doi.org/10.1073/pnas.1621129114); pmid: 28115689
- Z. Ding *et al.*, High-resolution cryo-EM structure of the proteasome in complex with ADP-AlFx. *Cell Res.* **27**, 373–385 (2017). doi: [10.1038/cr.2017.12](https://doi.org/10.1038/cr.2017.12); pmid: 28106073
- Y. Zhu *et al.*, Structural mechanism for nucleotide-driven remodeling of the AAA-ATPase unfoldase in the activated human 26S proteasome. *Nat. Commun.* **9**, 1360 (2018). doi: [10.1038/s41467-018-03785-w](https://doi.org/10.1038/s41467-018-03785-w); pmid: 29636472
- F. Beck *et al.*, Near-atomic resolution structural model of the yeast 26S proteasome. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 14870–14875 (2012). doi: [10.1073/pnas.1213333109](https://doi.org/10.1073/pnas.1213333109); pmid: 22927375
- A. Henderson, J. Eroles, M. A. Hoyt, P. Coffino, Dependence of proteasome processing rate on substrate unfolding. *J. Biol. Chem.* **286**, 17495–17502 (2011). doi: [10.1074/jbc.M110.212027](https://doi.org/10.1074/jbc.M110.212027); pmid: 21454622
- E. J. Worden, K. C. Dong, A. Martin, An AAA Motor-Driven Mechanical Switch in Rpn11 Controls Deubiquitination at the 26S Proteasome. *Mol. Cell* **67**, 799–811.e8 (2017). doi: [10.1016/j.molcel.2017.07.023](https://doi.org/10.1016/j.molcel.2017.07.023); pmid: 28844860
- M. Sen *et al.*, The ClpXP protease unfolds substrates using a constant rate of pulling but different gears. *Cell* **155**, 636–646 (2013). doi: [10.1016/j.cell.2013.09.022](https://doi.org/10.1016/j.cell.2013.09.022); pmid: 24243020
- O. Iosefson, A. R. Nager, T. A. Baker, R. T. Sauer, Coordinated gripping of substrate by subunits of a AAA+ proteolytic machine. *Nat. Chem. Biol.* **11**, 201–206 (2015). doi: [10.1038/nchembio.1732](https://doi.org/10.1038/nchembio.1732); pmid: 25599533
- O. Iosefson, A. O. Olivares, T. A. Baker, R. T. Sauer, Dissection of Axial-Pore Loop Function During Unfolding and Translocation by a AAA+ Proteolytic Machine. *Cell Rep.* **12**, 1032–1041 (2015). doi: [10.1016/j.celrep.2015.07.007](https://doi.org/10.1016/j.celrep.2015.07.007); pmid: 26235618
- F. Zhang *et al.*, Structural insights into the regulatory particle of the proteasome from *Methanocaldococcus jannaschii*. *Mol. Cell* **34**, 473–484 (2009). doi: [10.1016/j.molcel.2009.04.021](https://doi.org/10.1016/j.molcel.2009.04.021); pmid: 19481527
- D. M. Rubin, M. H. Glickman, C. N. Larsen, S. Dhruvakumar, D. Finley, Active site mutants in the six regulatory particle

- ATPases reveal multiple roles for ATP in the proteasome. *EMBO J*. **17**, 4909–4919 (1998). doi: [10.1093/embj/17.17.4909](https://doi.org/10.1093/embj/17.17.4909); pmid: [9724628](https://pubmed.ncbi.nlm.nih.gov/9724628/)
44. Y. C. Kim, X. Li, D. Thompson, G. N. Demartino, ATP-binding by proteasomal ATPases regulates cellular assembly and substrate-induced functions of the 26S proteasome. *J. Biol. Chem.* **288**, 3334–3345 (2013). doi: [10.1074/jbc.M112.424788](https://doi.org/10.1074/jbc.M112.424788)
45. H. Han, N. Monroe, W. I. Sundquist, P. S. Shen, C. P. Hill, The AAA ATPase Vps4 binds ESCRT-III substrates through a repeating array of dipeptide-binding pockets. *eLife* **6**, e31324 (2017). doi: [10.7554/eLife.31324](https://doi.org/10.7554/eLife.31324); pmid: [29165244](https://pubmed.ncbi.nlm.nih.gov/29165244/)
46. R. A. Maillard *et al.*, ClpX(P) generates mechanical force to unfold and translocate its protein substrates. *Cell* **145**, 459–469 (2011). doi: [10.1016/j.cell.2011.04.010](https://doi.org/10.1016/j.cell.2011.04.010); pmid: [21529717](https://pubmed.ncbi.nlm.nih.gov/21529717/)
47. P. Rodriguez-Aliaga, L. Ramirez, F. Kim, C. Bustamante, A. Martin, Substrate-translocating loops regulate mechanochemical coupling and power production in AAA+ protease ClpXP. *Nat. Struct. Mol. Biol.* **23**, 974–981 (2016). doi: [10.1038/nsmb.3298](https://doi.org/10.1038/nsmb.3298); pmid: [27669037](https://pubmed.ncbi.nlm.nih.gov/27669037/)
48. T. Sone, Y. Saeki, A. Toh-e, H. Yokosawa, Sem1p is a novel subunit of the 26 S proteasome from *Saccharomyces cerevisiae*. *J. Biol. Chem.* **279**, 28807–28816 (2004). doi: [10.1074/jbc.M403165200](https://doi.org/10.1074/jbc.M403165200); pmid: [15117943](https://pubmed.ncbi.nlm.nih.gov/15117943/)
49. D. S. Leggett, M. H. Glickman, D. Finley, Purification of proteasomes, proteasome subcomplexes, and proteasome-associated proteins from budding yeast. *Methods Mol. Biol.* **301**, 57–70 (2005). pmid: [15917626](https://pubmed.ncbi.nlm.nih.gov/15917626/)
50. H. C. Kim, J. M. Huijbreghse, Polyubiquitination by HECT E3s and the determinants of chain type specificity. *Mol. Cell. Biol.* **29**, 3307–3318 (2009). doi: [10.1128/MCB.00240-09](https://doi.org/10.1128/MCB.00240-09); pmid: [19364824](https://pubmed.ncbi.nlm.nih.gov/19364824/)
51. J. G. Narby, Coupled assay of Na⁺/K⁺-ATPase activity. *Methods Enzymol.* **156**, 116–119 (1988). doi: [10.1016/0076-6879\(88\)56014-7](https://doi.org/10.1016/0076-6879(88)56014-7); pmid: [2835597](https://pubmed.ncbi.nlm.nih.gov/2835597/)
52. B. Carragher *et al.*, Legion: An automated system for acquisition of images from vitreous ice specimens. *J. Struct. Biol.* **132**, 33–45 (2000). doi: [10.1006/jsbi.2000.4314](https://doi.org/10.1006/jsbi.2000.4314); pmid: [11121305](https://pubmed.ncbi.nlm.nih.gov/11121305/)
53. G. C. Lander *et al.*, Appion: An integrated, database-driven pipeline to facilitate EM image processing. *J. Struct. Biol.* **166**, 95–102 (2009). doi: [10.1016/j.jsb.2009.01.002](https://doi.org/10.1016/j.jsb.2009.01.002); pmid: [19263523](https://pubmed.ncbi.nlm.nih.gov/19263523/)
54. S. Q. Zheng *et al.*, MotionCor2: Anisotropic correction of beam-induced motion for improved cryo-electron microscopy. *Nat. Methods* **14**, 331–332 (2017). doi: [10.1038/nmeth.4193](https://doi.org/10.1038/nmeth.4193); pmid: [28250466](https://pubmed.ncbi.nlm.nih.gov/28250466/)
55. K. Zhang, Gctf: Real-time CTF determination and correction. *J. Struct. Biol.* **193**, 1–12 (2016). doi: [10.1016/j.jsb.2015.11.003](https://doi.org/10.1016/j.jsb.2015.11.003); pmid: [26592709](https://pubmed.ncbi.nlm.nih.gov/26592709/)
56. S. H. Scheres, S. Chen, Prevention of overfitting in cryo-EM structure determination. *Nat. Methods* **9**, 853–854 (2012). doi: [10.1038/nmeth.2115](https://doi.org/10.1038/nmeth.2115); pmid: [22842542](https://pubmed.ncbi.nlm.nih.gov/22842542/)
57. D. K. Manian, B. O. Forsberg, S. H. Scheres, E. Lindahl, Accelerated cryo-EM structure determination with parallelisation using GPUs in RELION-2. *eLife* **5**, e18722 (2016). doi: [10.7554/eLife.18722](https://doi.org/10.7554/eLife.18722); pmid: [27845625](https://pubmed.ncbi.nlm.nih.gov/27845625/)
58. E. F. Pettersen *et al.*, UCSF Chimera—a visualization system for exploratory research and analysis. *J. Comput. Chem.* **25**, 1605–1612 (2004). doi: [10.1002/jcc.20084](https://doi.org/10.1002/jcc.20084); pmid: [15264254](https://pubmed.ncbi.nlm.nih.gov/15264254/)
59. P. V. Afonine *et al.*, Towards automated crystallographic structure refinement with phenix.refine. *Acta Crystallogr. D Biol. Crystallogr.* **68**, 352–367 (2012). doi: [10.1107/S0907444912001308](https://doi.org/10.1107/S0907444912001308); pmid: [22505256](https://pubmed.ncbi.nlm.nih.gov/22505256/)
60. P. Emsley, B. Lohkamp, W. G. Scott, K. Cowtan, Features and development of Coot. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 486–501 (2010). doi: [10.1107/S0907444910007493](https://doi.org/10.1107/S0907444910007493); pmid: [20383002](https://pubmed.ncbi.nlm.nih.gov/20383002/)
61. V. B. Chen *et al.*, MolProbity: All-atom structure validation for macromolecular crystallography. *Acta Crystallogr. D Biol. Crystallogr.* **66**, 12–21 (2010). doi: [10.1107/S0907444909042073](https://doi.org/10.1107/S0907444909042073); pmid: [20057044](https://pubmed.ncbi.nlm.nih.gov/20057044/)
62. T. D. Goddard *et al.*, UCSF ChimeraX: Meeting modern challenges in visualization and analysis. *Protein Sci.* **27**, 14–25 (2018). doi: [10.1002/pro.3235](https://doi.org/10.1002/pro.3235); pmid: [28710774](https://pubmed.ncbi.nlm.nih.gov/28710774/)

ACKNOWLEDGMENTS

We thank B. M. Gardner, J. A. M. Bard, A. L. Yokom, and C. Puchades for comments and critical evaluation of the manuscript; all members of the Martin and Lander laboratories

for discussions, suggestions, and support; and R. J. Beckwith for cloning the model substrate. All cryo-EM data were collected at The Scripps Research Institute (TSRI) electron microscopy facility. We thank B. Anderson for microscope support and J. C. Ducom at TSRI's High Performance Computing facility for computational support. **Funding:** A.H.P. is a fellow of the American Cancer Society (132279-PF-18-189-01-DMC). S.N.G. is a Howard Hughes Medical Institute Fellow of the Damon Runyon Cancer Research Foundation (DRG-2342-18). G.C.L. is supported as a Pew Scholar in the Biomedical Sciences by the Pew Charitable Trusts. A.M. is an investigator of the Howard Hughes Medical Institute. This work was funded by the National Institutes of Health (DP2EB020402 to G.C.L. and R01-GM094497 to A.M.) and the Howard Hughes Medical Institute (A.M.). Computational analyses of EM data were performed using shared instrumentation at SR funded by NIH S100D021634.

Author contributions: A.H.P., E.A.G., and S.N.G. designed experiments and performed data analysis. A.H.P. performed cryo-EM sample preparation, data collection, and data processing. E.A.G. expressed and purified constructs and performed biochemical experiments. G.C.L. and A.M. designed and supervised the study. All authors wrote the manuscript.

Competing interests: The authors declare no competing interests. **Data and materials availability:** Structural data are available in the Electron Microscopy Databank and the RCSB Protein Databank (EMDB IDs 9042, 9043, 9044, and 9045 and PDB IDs 6EF0, 6EF1, 6EF2, and 6EF3 for the proteasome states 1D*, 5D, 5T, and 4D, respectively). All other data are available in the main text or the supplementary materials.

SUPPLEMENTARY MATERIALS

www.sciencemag.org/content/362/6418/eaav0725/suppl/DC1
Figs. S1 to S7
Tables S1 to S2
Reference (63)
Movies S1 and S2

10 August 2018; accepted 4 October 2018
Published online 30 October 2018
[10.1126/science.aav0725](https://doi.org/10.1126/science.aav0725)

Substrate-engaged 26S proteasome structures reveal mechanisms for ATP-hydrolysis–driven translocation

Andres H. de la Peña, Ellen A. Goodall, Stephanie N. Gates, Gabriel C. Lander and Andreas Martin

Science **362** (6418), eaav0725.

DOI: 10.1126/science.aav0725 originally published online October 11, 2018

Molecular-motor coordination

The proteasome is a cytosolic molecular machine that recognizes and degrades unneeded or damaged proteins that have been tagged with ubiquitin. A heterohexameric adenosine triphosphatase motor pulls the substrate into the proteolytic chamber, while at the same time, a protein located at the entrance of this motor removes the ubiquitin. De la Peña *et al.* trapped the substrate inside the motor by inhibiting removal of ubiquitin. This allowed them to determine cryo-electron microscopy structures in the presence of substrate and adenosine triphosphate (ATP). The findings distinguish three sequential conformational states that show how ATP binding, hydrolysis, and phosphate release are coordinated between the six subunits of the motor to cause the conformational changes that translocate the substrate through the proteasome.

Science, this issue p. eaav0725

ARTICLE TOOLS

<http://science.sciencemag.org/content/362/6418/eaav0725>

SUPPLEMENTARY MATERIALS

<http://science.sciencemag.org/content/suppl/2018/10/10/science.aav0725.DC1>

REFERENCES

This article cites 63 articles, 16 of which you can access for free
<http://science.sciencemag.org/content/362/6418/eaav0725#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)